

**Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA) concentrations in serum of renal failure patients by ELISA.**

**Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA) concentrations in serum of renal failure patients by ELISA.**

**\*\*Thekra A. Humada ,Ph.D medical microbiology ,Medicine College -**

**Tikrituniversity ,Assistant Professor**

**\*\*Hind I .Abdullah ,M.Sc medical microbiology ,Dentistry College -Tikrit**

**university ,Assistant Lecturer**

**Abstract**

*P.aeruginosa* is one of the most danger cause of morbidity and mortality in patients in nosocomial infection and renal failure. ELISA method this assay has high sensitivity and excellent specificity for detection of human PEA. The medical records of 64 patients (42 males and 22 females of renal failure patients ) undergoing hemodialysis at the Department of Nephrology–hemodialysis of the Tikrit Teaching Hospital from (12 - 2012 to 12-2013 ) were retrospectively reviewed. Control of exotoxin concentration is zero because the minimum detectable dose of human PEA concentration is typically less than 0.039ng/ml depend on sensitivity of ELISA kit. Detectable exotoxin A in serum of 64 patients The result show in this study maximum concentration was 3.5 ng/ml .positive result was 37 males of 42 and 17 females of 22. and no significant exotoxin A concentration between males and females.

**Key word:** *P.aeruginosa* , Renal failure , Exotoxin A ,Patients ,Serum ,ELISA.

**INTRODUCTION**

Patients with end-stage renal disease requiring dialysis are at increased risk for bloodstream infection ,this type of infection represents a main cause of morbidity and cause death. (1,2,3,4) . Virulence of *P. aeruginosa* is multifactorial and has been attributed to cell associated factors like exotoxin A (5,6). pathogenic *p. aureoginosa* strains possess a type III secretion system that allow them to deliver toxins directly into the cytoplasm of a host cell. Exotoxin A which causes tissue necrosis since it block protein synthesis(7) . The most important factor in the pathogenicity of *P. aeruginosa*, ETA consists of two subunits; fragment A is catalytic, and fragment B is responsible for interaction with eukaryotic cell receptors. ETA is cytotoxic to numerous mammalian cells tubular necrosis of kidneys.(8,9,10)

**MATERIALS AND METHODS**

**Patients :** The medical records of 64 patients (42 males and 22 females of renal failure patients ) undergoing hemodialysis at the Department of Nephrology–hemodialysis of the Tikrit Teaching Hospital from (12 -2012 to 12-2013 ) were retrospectively reviewed.

Control of exotoxin concentration is zero because the minimum detectable dose of human PEA concentration is typically less than 0.039ng/ml depend on sensitivity of ELISA kit ,that kit use in this method [human *pseudomonas* exotoxin A (PEA) ELISA Kit / Cusabio- china]



## Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA) concentrations in serum of renal failure patients by ELISA.

The free EDTA tubes (non-EDTA blood) was left about 15 min. in room temperature, the blood clot was detached from tubes surface sides to free the clot and let serum to accumulate at the clot surface. Centrifuged at 400 rpm (18 C°) for 10 min., the supernatant (serum was aspirated by Pasteur pipette, and re-centrifuged the supernatant in the same manner to sediment any erythrocytes may cause serum contamination, the supernatant was aspirated, store at -20 C° in tubes labeled with patient's number.

### ❖ Human pseudomonas exotoxin A (PEA) ELISA Kit

- Principle of the assay : This assay employs the quantitative sandwich enzyme immunoassay technique. Antibody specific for PEA has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any PEA present is bound by the immobilized antibody. After removing any unbound substances, a biotin-conjugated antibody specific for PEA is added to the wells. After washing, avidin conjugated Horseradish Peroxidase (HRP) is added to the wells. Following a wash to remove any unbound avidin-enzyme reagent, a substrate solution is added to the wells and color develops in proportion to the amount of PEA bound in the initial step. The color development is stopped and the intensity of the color is measured.

- Procedure :Kit were prepared as instructed by manufacturing Companies.

### **CALCULATION OF RESULTS**

Using the professional soft "Curve Expert 1.4" to make a standard curve is recommended, which can be downloaded from Cusabio web site.

### **Results**

PEA concentrations in 64 renal failure patients were estimated in serum by using ELISA .

Table (2) , show the mean concentration of exotoxin A of *Pseudomonas* for 42 males and 22 females of renal failure patients . there was no significant in exotoxin A concentration between males and females . Control of exotoxin concentration is zero because the minimum detectable dose of human PEA concentration is typically less than 0.039ng/ml depend on sensitivity of ELISA kit ,that kit use in this method [human *Pseudomonas* exotoxin A (PEA) ELISA Kit / Cusabio- china]

Figure (2) show detectable exotoxin A in serum of 64 patients .The result showin our study maximum concentration was 3.5 ng/ml .

Rational Function:  $y=(a+bx)/(1+cx+dx^2)$   
(manual of kit )

Coefficient Data:

a = -0.2258472

b = 1.4496171

c = -0.10786801

d = -0.043816625

X = OD(absorbtion)

Y = Concentration of exotoxin A

Also , in our study show no significant exotoxin A between males and females who have toxin in serum. Show in table (3) positive result was 37 males of 42 and 17 females of 22.

### **Discussion**

In this study , show the result of 64 renal failure patients (37 males and 17 females )were



**Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA) concentrations in serum of renal failure patients by ELISA.**

have exotoxin A (PEA) concentration average from (0.039 to 3.5) ng/ml in serum produced by causative agent *Pseudomonas aeruginosa*. Saroj Sharma, Ramanjeet Kaur, Vanashree Yadav, Kusum Harjai and Kusum Joshi.2004 show the importance of this organism is of special relevance since it is UTIs third leading cause, accounting for about 11% of nosocomial UTIs. In the present study, an exotoxin A-producing strain of *P. aeruginosa* PAO, and its mutant lacking this ability were employed to study the possible role of exotoxin A in acute as well as in chronic pyelonephritis.(11)

*P. aeruginosa* exotoxin A (toxin, lethal toxin), which is produced by over 90% of *P. aeruginosa* clinical isolates, However, some strains of *Pseudomonas* do not produce good yields of exotoxin A when grown in chemically defined medium (12)

Our study the first attempted to evolution exotoxin A in human serum, all previous studies on serum of laboratory animals, or determinate exotoxin A in broth media and most of studies determinate antibodies in serum against exotoxin A(13,14)and show in this result no different in exotoxin concentration between males or females. This result agreed with the pathogenic substances of *Pseudomonas aeruginosa* include chemoattractant factor,(15) numerous exoproducts such as exotoxins, proteases, phospholipase and leucocidin. In disease states such as cystic fibrosis, many clinical isolates are positive for alkaline protease, elastase and exotoxin A production (16,17). These enzymes have been implicated as important factors contributing directly or indirectly to the pathogenicity of microbes. Patients with UTI and kidney disease have a high prevalence of *Pseudomonas aeruginosa* colonization(18,19), which rapidly causes a chronic infection of the mucosal surface of the. This was also tested for exotoxin A by using two culture

supernatants at concentrations of 5.4 and 10.8 µg/l for different strains. Also, agree with the study. Acute *Pseudomonas aeruginosa* was established in guinea pigs by intratracheal instillation of bacteria. Challenge strains included PAO-1, a strain known to produce exotoxin A, alkaline protease, and elastase, and several PAO-1 mutants deficient in either biologically active exotoxin A or elastase production. Survival, intraperitoneal killing of bacteria, and blood cultures were compared among the groups. Strains of *P. aeruginosa* deficient in active elastase production appeared to be less virulent than the parent strain and were more easily cleared from the lung. Opposite results were obtained for the exotoxin A-deficient mutants. These data suggest that elastase, but not exotoxin A, was an important virulence factor during acute disease due to *P. aeruginosa*. Experimental data show that elastase and exotoxin A elicit high levels of antibodies both in experimental animals and in patients. These results suggest that these proteins should be considered for use in a prophylactic *Pseudomonas* vaccine.(20,21,22)

### **References**

- 1-Maria Fysaraki, George Samonis, Antonis Valachis, Eugenios Daphnis, Drosos E. Karageorgopoulos, Matthew E. Falagas, Kostas Stylianou, Diamantis P. Kofteridis, Incidence, Clinical, Microbiological Features and Outcome of Bloodstream Infections in Patients Undergoing Hemodialysis. *International Journal of Medical Sciences*, 2013. 10(12):1632-1638. doi: 10.7150/ijms.6710
- 2.Tozawa M, Iseki K, Fukiyama K. Prevalence of hospitalization and prognosis of patients on chronic dialysis. *Clin Exp Nephrol* L 2000;4:236-240



**Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA)  
concentrations in serum of renal failure patients by ELISA.**

3. US Renal Data System, The National Institutes of Health, National Institute of Diabetes, Digestive and Kidney Diseases, Bethesda, MD). USRDS 2008 Annual Data Report.
4. Liu JW, Su YK, Liu CP, et al. Nosocomial blood-stream infections in patients with end-stage renal disease; excess length of hospital stay, extracost and attributed mortality. *Hosp Infect*; 2002, 50:224-227
5. Matheson NR, Potempa J, Travis J. Interaction of a novel form of *Pseudomonas aeruginosa* alkaline protease (aeruginolysin) with interleukin-6 and interleukin-8. *Biol Chem*; 2006, 387:911-5.
6. Zulianello L, Canard C, Kohler T, Caille D, Lacroix JS, Meda P. Rhamnolipids are virulence factors that promote early infiltration of primary human airway epithelia by *Pseudomonas*, 2000.
7. Al-Rubaiee, L., "The Role of *Pseudomonas aeruginosa* in chronic suppurative otitis media infection". Thesis M.Sc., Medicine collage, University of Baghdad, 2009, pp.5, 6, 20, 22.
8. Pinghui, V. Liu. Extracellular toxins of *Pseudomonas aeruginosa*. *The journal of infection and diseases*, 1974, 130: 594-599.
9. Woods, D. E., and B. H. Iglewski. Toxins of *Pseudomonas aeruginosa*: new perspectives. *Rev. Infect. Dis.* 1983, 5:714- 722.
10. Middlebrook, J. L, and R. B. Dorland. Response of cultured mammalian cells to the exotoxins of *Pseudomonas aeruginosa* and *Corynebacterium diphtheriae*: differential cytotoxicity. *Can. J. Microbiol.* 1977, 23:183-189.
11. Saroj Sharma, Ramanjeet Kaur, Vanashree Yadav, Kusum Harjai and Kusum Joshi. Contribution of Exotoxin A of *Pseudomonas aeruginosa* in Acute and Chronic Experimental Renal Infection. *Jpn. J. Infect. Dis.*, 57, 119-120, 2004
12. Iglewski, B. H., and J. C. Sadoff. Toxin inhibitors of protein synthesis: production, purification and assay of *Pseudomonas aeruginosa* toxin A. *Methods Enzymol.* 1979, 60:780-793.
13. Moss, R. B., Hsu, Y.-P., Lewiston, N. J., Curd, J. G., Milgrom, H., Hart, S., Deyer, B. & Larrick, J. W. Association of systemic immune complexes, complement activation, and antibodies to *Pseudomonas aeruginosa* lipopolysaccharide and exotoxin A mortality in cystic fibrosis. *Am. Rev. Respir. Dis.* (1986), 733, 648-652.
14. Nicas, T. L & Iglewski, B. H. The contribution of exoproducts to virulence of *Pseudomonas aeruginosa*. *Can. J. Microbiol.* (1985), 37, 387-392.
15. Suter, S., Schaad, U. B., Roux, L., Nydegger, U. E. & Waldvogel, F. A. Granulocyte neutral proteases and *Pseudomonas aeruginosa* elastase as possible causes of airway damage in patients with cystic fibrosis. *J. Infect. Dis.* (1986), 149, 523-531.
16. Morihara, K. & Tsuzuki, H. Production of protease and elastase by *Pseudomonas aeruginosa* strains isolated from patients.

**Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA)  
concentrations in serum of renal failure patients by ELISA.**

Infect. Immun. (1977) ,75, 679-685.

17. Pollack, M., Taylor, N. S. & Callahan, L. T. Exotoxin production by clinical isolates of *Pseudomonas aeruginosa*.

Infect. Immun. (1977) . 75, 776-780.

18. Döring, G., Obernesser, H. J. & Botzenhart, K. Extracellular

toxins of *Pseudomonas aeruginosa*.

III. Radioimmunoassay for detection of alkaline protease. Zbl. Bakt.Hyg., I.

Abt. Orig. (1982) , A 252, 239-247.

19. Döring, G., Goldstein, W., Roll, A., Schiotz, P. O., Hoiby, N. &

Botzenhart, K. Role of *Pseudomonas aeruginosa* exoenzymes

in lung infections of patients with cystic fibrosis. Infect.

Immun. (1985) .49, 557-562.

20. Jaffar-Bandjee, M. C., Carrere, J., Lazdünski, A.; Guy-Crptte,

O. & Galäbert, C. Direct double antibody Sandwich

immunoassay for *Pseudomonas aeruginosa* elastase. J. Immunol.

Methods (1993) ,164, 27-32.

21. Liu, P. V. Exotoxins of *Pseudomonas aeruginosa*. I.

Factors that influence the production of exotoxin A. J. Infect.

Dis. (1973) , 725, 506-513.

22. Wilson, M. B. & Nakane, P. K. Recent developraents

in the periodate method of conjugate horseradish peroxidase

(HRPO) to antibodies. in: immunoßuorescence and Related

Staining Techniques (Knapp, W., ,Holubar, K. & Wick, G.,

eds.) Elsevier/North-Holland, Amsterdam, (1978) pp. 215-224.



**Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA) concentrations in serum of renal failure patients by ELISA.**

**Table(1)Materials provided in ELISA kit**

Quantity	Reagents
1(96 wells)	Assay plate (12 x 8 coated Microwells)
2	Standard (Freeze dried)
1 x 120 µl	Biotin-antibody (100 x concentrate)
1 x 120 µl	HRP-avidin (100 x concentrate)
1 x 15 ml	Biotin-antibody Diluent
1 x 15 ml	HRP-avidin Diluent
1 x 50 ml	Sample Diluent
1 x 20 ml	Wash Buffer (25 x concentrate)
1 x 10 ml	TMB Substrate
1 x 10 ml	Stop Solution
4	Adhesive Strip (for 96 wells)
1	Instruction manual

**sample: Serum Separation**

**Table (2) Toxin concentration in renal failure patient according to gender**

Gender	No.	Mean±SD
Male	42	0.814±0.95
Female	22	0.436±0.48

$t=1.744$ ,  $df=62$ ,  $P > 0.05$  not significant

**Table (3) The positive renal failure patient distribution according to serum toxin level and gender**

Toxin in serum	Male		Female	
	Frequency	Percent	Frequency	Percent
Positive (>0.039)ng/ml	37	88.1	17	77.3
Negative	5	11.9	5	22.7
Total	42	100	22	100

Yates' chi-square=0.593,  $df=1$ ,  $p > 0.05$  not significant

Quantitative determination of endogenic human *Pseudomonas aeruginosa* exotoxin A (PEA) concentrations in serum of renal failure patients by ELISA.

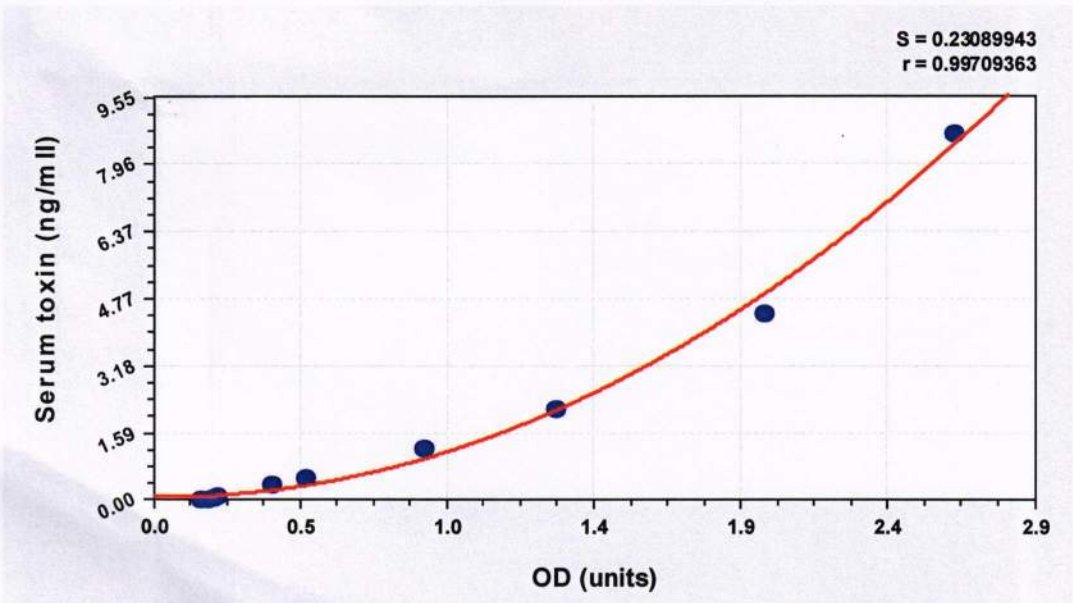


Figure (1): Standard curve of concentration *Pseudomonasaeruginosa*exotoxin A

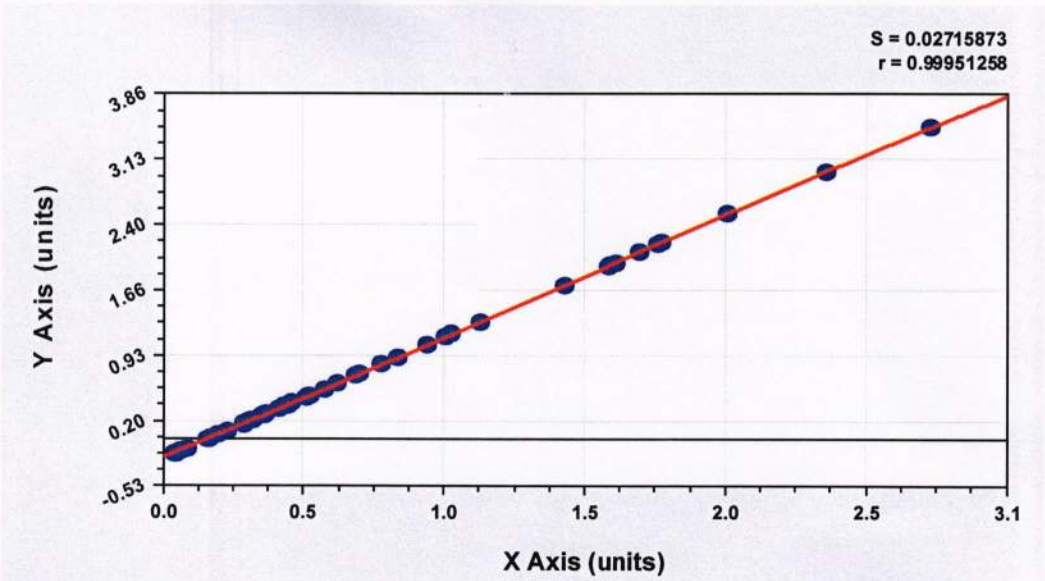


Figure (2) Quantitative determination of endogenic human *Pseudomonas* exotoxin A (PEA) concentrations in serum of renal failure patients by ELISA.